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Role of the Wnt signalling pathway in the development of endothelial disorders in response to hyperglycaemia

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Abstract

Introduction. Diabetes mellitus (DM) is the most common metabolic disease. A WHO report from 2016 indicates that 422 million people worldwide suffer from DM or hyperglycaemia because of impaired glucose metabolism. Chronic hyperglycaemia leads to micro- and macro-vessel damage, which may result in life-threatening complications. The Wnt pathway regulates cell proliferation and survival by modulating the expression of genes that control cell differentiation. Three linked Wnt pathways have been discovered thus far: a β -catenin-dependent pathway and two pathways independent of β -catenin – the planar cell polarity pathway and calcium-dependent pathway. The Wnt pathway regulates genes associated with inflammation, cell cycle, angiogenesis, fibrinolysis and other molecular processes.

Areas covered. This review presents the current state of knowledge regarding the contribution of the Wnt pathway to endothelial ageing under hyperglycaemic conditions and provides new insights into the molecular basis of diabetic endothelial dysfunction.

Conclusion. The β -catenin-dependent pathway is a potential target in the prophylaxis and treatment of early-stage diabetes-related vascular complications. However, the underlying molecular mechanisms remain largely undetermined and require further investigation.

Hyperglycaemia as the main cause of endothelial dysfunction and vascular ageing

The biological role of vascular endothelium

Endothelial cells (ECs) play a key role in the haemostasis and maintenance of vessel functions. ECs maintain a physical barrier between a vessel wall and the lumen. Additionally, the endothelial monolayer is a potent secretory tissue that releases a number of mediators that regulate several physiological processes, such as coagulation (Ref. 1), platelet aggregation (Refs 2, 3), fibrinolysis (Ref. 4) and vascular permeability (Ref. 5).

Under normal conditions, quiescent ECs maintain vessel homeostasis. The activation of ECs by pro-inflammatory cytokines, hypoxia or shear stress leads to a dramatic alteration in the EC phenotype. Activated ECs express larger amounts of growth factors, inflammatory mediators and adhesion molecules. The characteristic features of endothelial dysfunction are reduced endothelium-mediated vasorelaxation, impaired fibrinolytic ability and increased oxidant stress (Refs 6–10). At the cellular level, the disintegration of intercellular junctions formed by calcium-dependent adhesion molecules, VE-cadherins, is considered to be the most distinguished EC deterioration effect observed during endothelial dysfunction. This in turn leads to an increase in leukocyte adherence, rolling and subsequent extravasation (Refs 1, 9–12).

Endothelial hyperpermeability and microvessel leakage are thought to be crucial for the development of a variety of pathologies, including enhanced thrombotic processes and haemorrhagic shock, which are observed in diabetes mellitus (DM) (Refs 5, 9, 13).

Endothelial dysfunction as a preliminary stage of the ageing processes

During the life span of cells, irreversible ageing processes are physiological. In terms of 'endothelial ageing', the number of unfavourable changes in endothelium function begins to appear prominently and finally becomes irreversible, whereas the term 'endothelial dysfunction' refers to a loss of the physiological properties of the cell in a temporal manner. The intracellular and intercellular interactions that occur during 'endothelial ageing' appear to be crucial, and they can influence the microenvironment and alter cell signalling, including altered NO signalling, revasculating factor expression (integrin β 2, SDF-1) and release of endothelial progenitor cells (Ref. 14).

In physiological conditions, when the metabolic control is satisfactory, the blood glucose levels are maintained between 3.6 and 5.8 mmol/l. Human ECs are locally exposed to such glucose concentrations; thus, every recurrent (the short-term hyperglycaemia incubation (60–240 min) and long-term incubation(48 h)) increase in glucose level affects ECs with short- or long-term consequences (Refs 6, 15). Acute local hyperglycaemia (16.6 mmol/l) achieved by brachial intra-arterial infusion of 50% dextrose impairs endothelium-dependent vasodilatation in healthy subjects (Ref. 16). In normoglycaemia, the EC phenotype is described

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as quiescent (Ref. 6). The quiescence of these cells is closely controlled by several molecules, such as human Bone Morphogenetic Protein-9 (BMP-9) (Ref. 17), transcription factor E2-2 (Ref. 18), Angiopoietin 1 (Ang1) (Refs 19, 20) and very low-density lipoprotein (LDL) receptor (Ref. 21). The expression of the anti-angiogenic R-ras gene is a biomarker of vascular quiescence (Ref. 22).

Hyperglycaemia, in the context of ECs, is defined as exposure of the cells to glucose concentrations higher than 10 mmol/l (experiments carried out in vivo or in vitro) (Ref. 6). ECs cultured under hyperglycaemic conditions lose their quiescence and become dysfunctional (Ref. 6). Studies performed on vascular ECs cultured in hyperglycaemic conditions (30.5 mmol/l D-glucose) indicated that high glucose concentrations induce pro-adhesive and proinflammatory phenotypic changes (Refs 14, 23, 24). Under these conditions, the expression of interleukin (IL)-1 β and adhesion molecules, such as vascular cell adhesion molecule-1 (V-CAM1) or endothelial selectin (P-SEL), was upregulated (Ref. 24). Moreover, increased levels of IL-8, IL-6, vascular endothelial growth factor (VEGF), and tumour necrosis factor- α (TNF α) were also observed (Ref. 25).

Several cellular functions are regulated via interactions between cytoskeletal elements (actin filaments and microtubules) and adhesive molecules. The rearrangements of the EC cytoskeleton and adhesives change cell shape (Ref. 26). Targosz-Korecka *et al.* showed that long-lasting hyperglycaemic conditions result in cytoskeletal changes such as stress fibre formation and F-actin polymerisation. After 14 passages, cells cultured under hyperglycaemic conditions (25 mM D-glucose) exhibited a modification in the structure of the cortical actin cytoskeleton, which led to the formation of inter-cellular gaps and increased the permeability of the endothelium. A decrease in the integrity of the endothelial barrier and cohesiveness of the EC layer was observed after 20 passages (Ref. 27).

The nuclear factor Nrf2 is a key regulator in diabetes-induced endothelial dysfunction (Ref. 28). The duration of diabetes increases the expression of pro-inflammatory genes, such as chemokine ligands CCL2 and CCL5, which are controlled by Nrf2. The upregulation of mRNA levels for these chemokines was detected in aortic ECs in short-term diabetes induced in type 1 diabetic mice, whereas in the later stages of diabetes, upregulation was detected in the venous and aortic ECs. These observations suggest that during diabetes, endothelial dysfunction is induced both in arteries and veins to a different extent and in a different manner (Ref. 29).

Under physiological conditions, ECs show a low number of intracellular organelles, which are involved in degradation (lysosomes) or biosynthetic activities (Golgi apparatus and endoplasmic reticulum (ER)) and are produced in the basal lamina. Studies performed using human aortic ECs cultured under hyperglycaemic conditions and in vivo rodent studies have shown a significant enrichment in the number of biosynthetic organelles in these conditions (Refs 6, 30). The Golgi complex is observed in the capillary and aortic ECs, and interestingly, large volumes of rER have been observed in ECs of the athero-susceptible aortic arch, femoral artery and retinal venules. Direct and prolonged exposure to hyperglycaemia was associated with cytoskeleton reorganisation and mitochondrial fragmentation (Ref. 31). It is known that diabetes is linked to the thickening of the basal lamina of capillary ECs, resulting in a reduction in the number of metabolic products and nutrients transported between the tissue and circulation leading to endothelial dysfunction (Ref. 32).

Hyperglycaemia as a link to vascular ageing

Hyperglycaemia is one of the principal factors inducing endothelial dysfunction, the basis and hallmark of which are an increased synthesis of extracellular matrix (ECM) proteins, which plays a key role in the development of vascular complications in diabetic patients, such as retinopathy, nephropathy, hypertension and atherosclerosis (Refs 8, 14, 33, 34). Hyperglycaemia is known to modify the mechanical properties of ECs by increasing the stiffness of the cell membrane (Ref. 27).

A crucial phenomenon leading to ageing of ECs is the so-called 'glycaemic memory', which means the development of vascular complications, such as membrane stiffness or upregulation of cell adhesion molecules (ICAM and VCAM) (Ref. 27) after prolonged periods of hyperglycaemia despite a subsequent normalisation of glucose concentration. These phenotypic changes in ECs are controlled via epigenetic mechanisms, among which DNA methylation, histone modification and non-coding RNAs are considered important (Ref. 35).

Another key feature of EC ageing is an endothelial-tomesenchymal transition (EndoMT), a gradual process of phenotypic changes, from endothelium to mesenchymal cells. This process is observed when cells express lower levels of endothelial markers, such as vascular endothelial cadherin (VE-cad), claudin and cytokeratin-18, and lose their function along with higher expression of mesenchymal markers, such as vimentin, α -smooth muscle actin, and types I and III interstitial collagens, and acquire their function (Ref. 36). This process is regulated by transforming growth factor- β (TGF- β) family receptors and Wnt signalling pathways (β -catenin-dependent and -independent), which contribute to EndoMT in both vascular embryogenesis and cardiac tissue regeneration (Refs 36, 37). For example, a population of endocardial ECs that give rise to mesenchymal progenitors tend to migrate into the heart valves or differentiate into vSMCs and pericytes that cover the larger arteries in the heart (Ref. 38). A similar process has been observed in human aortic ECs exposed to glucose and in the hearts and kidneys of patients with DM (Refs 39, 40). Hyperglycaemia causes numerous modifications in the vascular tissue, which may accelerate atherosclerosis. Investigations of the vasculature of patients with diabetes and diabetic animals have shown three major mechanisms that are associated with pathological changes observed in these models: protein kinase C (PKC) activation, non-enzymatic glycosylation of proteins and lipids and oxidative stress. PKC is a key player in one of the Wnt signalling pathways, indicating that it could be involved in diabetes-augmented atherosclerosis (Ref. 41). Interestingly, in diabetic nephropathy (DN), exosomes derived from hyperglycaemic glomerular ECs have a significant impact on podocytes. Protecting glomerular ECs from undergoing EndoMT and inhibiting the release of TGF-β1-containing exosomes inhibits podocyte dysfunction and, therefore, renal fibrosis (Ref. 42).

Wnt signalling pathways

The Wnt signalling network has been studied for over 30 years, but it is still not completely understood (Refs 38, 43). The proteins involved in these pathways are evolutionarily highly conserved and present in a wide spectrum of organisms, ranging from marine anemones to humans, implying that Wnt pathways regulate fundamental cell processes. Indeed, Wnt signalling has been shown to regulate embryonic development through body axis patterning, cell specialisation, migration, polarisation, proliferation and differentiation (Ref. 44). Mutations in proteins of the Wnt network result in abnormalities of the bone (Ref. 45), heart and skeletal muscle structure (Ref. 46), and may lead to the development of cancer, DM (Ref. 47) or obesity (Ref. 48).

The key players that initiate the Wnt cascade include a large family of Wnt proteins, a family of transmembrane receptors, Fz (Frizzled), which is a group of co-receptors, the most important of which are lipoprotein receptor-related protein (LRP)-5/6,



Fig. 1. The β -catenin-dependent pathway in the absence of Wnt ligands. In the absence of Wnt ligands, DKK1 (Wnt antagonist) binds to the LRP5/6 receptor. β -catenin interacts with a degradation complex, which consists of APC, PP2A, AXIN, CK1 and GSK-3 β . β -catenin is phosphorylated by CK1 and GSK-3 β , which leads to the binding of β -TRCP to β -catenin and its ubiquitylation and degradation in proteasomes. Transcription of Wnt-responsive genes is blocked.

receptor tyrosine kinase (Ryk), and tyrosine-protein kinase transmembrane receptor (ROR2) (Figs 1 and 2). A crucial protein for the activation of the Wnt pathway is a cytoplasmic phosphoprotein Dishevelled (Dvl), which acts as a molecular switch to decide which of the Wnt pathways will be activated (Ref. 49). It has three domains, namely, DIX (amino-terminal), PDZ (central) and DEP (carboxy-terminal), and different combinations of these domains interact with each of the downstream Wnt pathways (Ref. 49). Although the current understanding of the aforementioned signalling is as a large network of mutual protein interactions, the division is still convenient for explanatory reasons.

The Wnt pathway is involved in the embryonic development of the pancreas (Ref. 50), insulin secretion (Ref. 51) and regulation of proliferation (Ref. 52) and survival of β -cells (Ref. 53). Experiments conducted on LRP receptor knockout mice showed

abnormalities in insulin secretion, leading to glucose intolerance (Ref. 54). A reduction in the expression of the TCF7L2 gene in human pancreatic islets resulted in an increase in the rate of apoptosis as well as a reduction in the level of phosphorylated Akt kinase (Refs 53, 55).

The Wnt pathway is active in human ECs (Ref. 55). Various Wnt signalling components, such as Fz receptors, Wnt ligands and effector TCF transcription factors, have been identified in ECs (Refs 56–58).

In vitro, sub-confluent ECs display more angiogenic characteristics than confluent ECs, with a specific TCF activity. In confluent cells, β -catenin was detected in intercellular gap junctions, whereas in sub-confluent cells, β -catenin localised in the nucleus and the cytoplasm. A diverse pattern of Wnt, Fzd, SFRP and Dkk gene expression in human ECs has also been



Fig. 2. The β -catenin-dependent pathway in the presence of Wnt ligands. Wnt ligands bind to the Frizzled receptor and LRP5/6. GSK-3 β and CK1 phosphorylate LRP5/6, which leads to the recruitment of Dvl and Axin to LRP5/6 and Frizzled. β -Catenin translocates to the nucleus, and after binding to LEF1/TCF, transcription factors induce Wnt-responsive gene transcription.

observed, which depends on the differentiation status of the cells (Ref. 59).

A possible mechanism for the development of endothelial disorders in diabetic conditions is EndoMT regulated by the crosstalk of the TGF- β and Wnt pathways (Ref. 36). Increased β -catenin expression enhances TGF- β sensitivity in hyperglycaemic conditions (Refs 38, 42). Unravelling these yet unclear relationships will have a great impact on our understanding of hyperglycaemia-induced endothelial dysfunction and vascular complications.

The β-catenin-independent Wnt pathways

The planar cell polarity pathway is independent of β -catenin (β -cat) and is activated by an interaction between one of the Wnt ligands and Fz. Similar to the β -catenin-dependent pathway,

Dvl is recruited, and its PDZ and DEP domains form a complex with Dishevelled-associated activator of morphogenesis 1 (DAAM1) (Fig. 3). This activates the Rho protein, and subsequently, the ROCK kinase – a protein responsible for cytoskeleton arrangement. Dvl also interacts with Rac1, which leads to actin polymerisation (via JNK) and the binding of profilin to actin filaments, which results in actin reorganisation and change in cell polarity (Ref. 60).

The Wnt/calcium pathway is the second β -cat-independent Wnt pathway. Upon Wnt-Fz interaction, the PDZ and DEP domains of Dvl are directly activated; Dvl also interacts with a G protein, resulting in the activation of either phospholipase C (PLC) or phosphodiesterase (PDE) (Refs 38, 61) (Fig. 4). An active PLC causes a rise in the levels of diacylglycerol (DAG) and inositol triphosphate (IP3) and subsequently results in the



Fig. 3. The PCP pathway in the presence of Wnt ligands. After binding of the Wnt ligand to the Frizzled receptor, Dvl and DAAM are activated. These two proteins activate the small GTPases Rho A and RAC-1, which leads to the activation of ROCK and JNK kinases. JNK phosphorylates JUN, which is translocated to the nucleus to regulate gene expression.

release of ionic calcium from the ER. This, *via* the activation of Cdc42, regulates cytoskeleton-dependent processes such as adhesion and migration of the cell. The increase in calcium level also activates CamKII (Ca2+/calmodulin-dependent protein kinase II), which activates kinases such as TAK1 (TGF- β -activated kinase I) and NLK (nemo-like kinase) that inhibit β -cat signalling in the β -catenin-dependent Wnt pathway (Ref. 49). On the other hand, the Dvl-G protein interaction can produce the opposite effect if PDE is activated. In this case, calcium release is not increased, and downstream processes do not occur (Ref. 60).

The β -catenin-dependent Wnt pathway

The involvement of β -cat distinguishes the β -catenin-dependent Wnt pathway from β -catenin-independent pathways. β -catenin

plays two crucial roles within the cell: it is part of the cadherin anchoring system mediating cell-cell interactions and acts as a transcription co-activator in the cell nucleus. In the absence of Wnt ligands, excess β -cat is degraded by a protein complex consisting of Axin, adenomatous polyposis coli (APC), protein phosphatase 2A (PP2A), glycogen synthase kinase 3 (GSK3) and casein kinase 1 α (CK1 α) (Fig. 1) (Ref. 62). Binding of a Wnt ligand to Fz and a simultaneous activation of the LRP 5/6 co-receptor recruits Axin to the plasma membrane, resulting in the disassembly of the degradation complex. Moreover, activated Dvl inactivates GSK-3. Owing to the inactivation of Axin and GSK-3, β -cat is not destroyed and is subsequently transported into the nucleus, where it acts as a co-activator for the TCF/ LEF transcription factors and activates the target gene expression (Fig. 2) (Ref. 63). Target genes for β -cat-TCF/LEF appear to be



Fig. 4. The β-catenin-independent pathway in the presence of Wnt ligands. After binding of the Wnt ligand to the Frizzled receptor, the Dvl protein activates PLC, leading to the cleavage of PtdIns(4,5)P into DAG and InsP3. InsP3 increases the cytoplasmic free-calcium level, which subsequently activates CAMKII, PKC and calcineurin. Calcineurin activates NFAT, which is translocated into the nucleus and activates the transcription of Wnt-responsive genes.

specific for the cell type. No universal target genes have been identified thus far. One of the most plausible candidates may be Axin2/conductin (Ref. 64), which is involved in the autoregulation of the pathway. For an extensive, updated list of Wnt target genes, the following website '*the Wntwebpage*' is recommended http://web.stanford.edu/group/nusselab/cgi-bin/wnt/.

Experiments carried out on two macrophage cell lines (RAW264.7 and J774.2) indicate that glucose concentration regulates β -catenin levels by promoting autocrine activation of the β -catenin-dependent pathway. This activation is mediated by changes in N-linked glycosylation of proteins and the hexosamine pathway (Ref. 60). The complexity of the catenin-dependent Wnt pathway in cell reprogramming in preadipose cells, where the Wnt antagonist Dickkopf1 (Dkk1) and its receptors (Krm1, LRP5 and LRP6), is coordinately regulated during the early stages

of human adipogenesis in vitro (Ref. 65). Crosstalk between cytokine and Wnt signalling networks has also been reported; for example, TNF- α inhibits adipogenesis via a β -catenin/TCF4 (TCF7L2)-dependent pathway, which suggests a possible mechanism in endothelial dysfunction (Ref. 66).

The effects of β-catenin-dependent Wnt signalling

High glucose concentrations disrupt endothelial adherent junctions (AJs) by dissociating the VE-cad- β -cat complex via activation of the Wnt/ β -cat pathway. This process is mediated by tyrosine phosphorylation of VE-cad through PKC- β and MLC phosphorylation (Ref. 67). Blocking the E1E2 domain of Wnt co-receptor LRP6 attenuates the accumulation of β -cat and overexpression of VEGF, intercellular adhesion molecule-1 (ICAM-1),



Fig. 5. TERT and β -catenin interaction. (A) During cell division, the telomerase complex repairs the chromosome ends in progenitor cells. TERT ensures reverse transcriptase activity to this complex and uses TERC as a template. (B) TERT increases the transcriptional activity of the β -catenin/TCF complex via interaction with BRG1. Both functions of TERT might increase the proliferation of progenitor cells and prevent cellular senescence at the same time.

and TNF- α induced by high glucose concentrations in retinal ECs (Ref. 68). This suggests that antibodies blocking β -catenindependent Wnt signalling can protect against vascular leakage and inflammation in the retina of DR models, which may be used as therapeutic agents in combination with other antiangiogenic compounds. The possible molecular mechanism underlying this is the regulation of the β -cat and TCF/LEF target genes, including myc and cycD1, which promote vascular cell survival, proliferation and migration (Refs 69–71).

Another important molecule linking Wnt signalling to cell longevity is a protein component of the telomerase complex, telomerase reverse transcriptase (TERT), which provides the reverse transcriptase activity of telomerase and is responsible for DNA synthesis at the ends of chromosomes. TERT is essential for maintaining genome stability and cell immortality. High telomerase activity is detected in many human cancers, whereas it is maintained at a low level in somatic cells (Ref. 72). Since uncontrolled cell viability underlies a number of diabetes-related conditions, such as atherosclerosis, DR and DN, regulating telomerase activity is an attractive approach in the development of novel therapeutic strategies for these diseases.

c-Myc, Sp1 (specificity protein 1), AP2 (activator protein 2) and HIF-1 (hypoxia-inducible factor 1) are known regulators of human TERT in normal and cancer cells (Refs 73, 74). Wnt pathways have been found to regulate TERT (Fig. 5). FH535, a β -cat/ TCF complex inhibitor, significantly inhibits telomerase activity in many cell lines, such as HCT116, MCF7 and MCF10A (Ref. 75). The results of the same study showed that activation of the Wnt pathway stimulates hTERT expression and telomerase activity. In this context, an interesting novel molecule is BRG1, which is a subunit of a protein complex responsible for alterations in chromatin conformation, facilitating transcription. BRG1 has been shown to be a component of TERT protein complexes. Binding of β -cat to BRG1 results in stimulation of β -cat target gene expression. Therefore, the BRG1-TERT complex might be a molecular link between β -cat and TERT. Experiments carried out on cells from the murine small intestine showed that a complex composed of β -cat, TERT and TCF protein bound to β -cat target genes, whereas the deletion of TERT decreased the expression of Wnt target genes in mouse embryonic stem cells (Ref. 76).

Hyperglycaemia and Wnt signalling in diabetes-associated neoplasms

Some types of cancers are known to be more abundant in diabetic and obese individuals (Ref. 77). This phenomenon has been partially attributed to the growth factor activity of insulin, which is present in higher concentrations in obese patients with insulin resistance or early stages of diabetes. However, in 2013, Chocarro-Calvo et al. (Ref. 78) showed that glucose is essential for β -catenin accumulation in the nucleus. Following Wnt activation, β -catenin accumulates in the cytoplasm, but in the absence of glucose, it is not transported to the nucleus. Upon the addition of glucose, β -catenin is rapidly translocated to the nucleus, where it forms a complex with LEF1, which functions as a p300 acetylase complex and simultaneously reduces the activity of SIRT1 deacetylase. This leads to acetylation of β -catenin, its accumulation in the nucleus, and subsequent activation of target genes. It needs to be emphasised that glucose alone, without a Wnt ligand, did not inactivate GSK-3 β . Moreover, hyperglycaemia has been shown to be a potent amplifier of β -cat/Wnt activity in cancer cell lines associated with diabetes and hyperglycaemic conditions. In hepatocellular carcinoma, high glucose concentrations activate β -catenin-dependent Wnt signalling, which is mediated by DKK4 suppression and enhanced the β -cat activity, leading to loss of check at G0/G1/S phases (Ref. 79). This may partly explain the relationship between hyperglycaemia and tumour progression.

A relevant role of DM has been observed in the development of solid organ malignancies, for example, pancreatic (Refs 80, 81), breast (Refs 82, 83), liver (Refs 83, 84), endometrial (Refs 85–87), colorectal (Refs 80, 88) and bladder cancers (Refs 89, 90). The strongest association with liver and pancreatic cancer was observed in patients with DM2. Bearing in mind that hyperinsulinaemia is often associated with hyperglycaemia, some experiments were performed on cancer cell lines cultured in high glucose (11 mM) and insulin (100 ng/ml) concentrations. Both conditions caused increased proliferation of cancer cell lines such as SW480 (human colorectal carcinoma), MCF-7 (human breast adenocarcinoma), HT29 (human colon carcinoma), MDA MB468 (human breast adenocarcinoma), PC3 (human prostate cancer) and T24 (human bladder carcinoma) (Ref. 91).

Large population studies show that there is a different pattern for the risk of incidence and mortality of a number of cancer types in T1DM and T2DM individuals. T1DM is associated with a higher incidence of pancreas, liver, oesophagus, colon and rectum. Moreover, for women with T1DM, a higher risk of stomach, thyroid, brain, lung, endometrium and ovary cancers has been reported. In T2DM, almost all site-specific cancers are more likely to develop, with the highest risk recorded for liver and pancreas. Given the different pathogenesis of T1DM and T2DM, with hyperinsulinaemia present only in the latter, it seems that hyperglycaemia might be the dominant trigger eventually leading to alterations in epigenetic regulation (Refs 78, 79). The pro-inflammatory role of insulin signalling has been observed in the development and progression of different types of cancers in both types of DM (Refs 80, 83-85). Interestingly, there seems to be a protective role of both types of DM with regard to prostate cancer incidence. For T2DM, when co-existing obesity is very frequent, this has been justified by obesity-related lower testosterone levels; however, this does not explain the relationship with T1DM (Ref. 92).

Wnt signalling and atherogenesis in hyperglycaemic conditions

Atherosclerosis is common among patients with DM. It is marked by the appearance of plaques consisting of LDL cholesterol, leukocytes, smooth muscle cells and lipids in the arterial walls (Ref. 93). The formation of an atherosclerotic plaque is a complex process, and the plaques are more frequently found at branching points within the vasculature, coinciding with turbulent blood flow, rather than the straight sections where the luminal ECs undergo lower shear stress. As described before, the activated and inflamed endothelium loses its protective abilities as it undergoes EndoMT and acts as a source of osteogenic progenitors, leading to vascular calcification and allowing enhanced transendothelial migration of leukocytes (Ref. 36). There is growing evidence that the Wnt pathway is also an important part of the pathophysiology of these processes. Wnt5A acts via the Ca2+-dependent pathway and has been shown to induce the expression of pro-inflammatory cyclooxygenase-2 in ECs (Ref. 94). Wnt5A has also been observed to be abundant in macrophages within atherosclerotic plaques (Refs 69, 95). The expression of Wnt5A mRNA is activated by exposure to ox-LDL in human macrophages (Ref. 96). Serum concentrations of Wnt-5A were higher in patients with advanced atherosclerosis than in healthy individuals (Ref. 97). Moreover, Wnt signalling appears to be an important part of the pathophysiology of myocardial infarction (MI) and its complications. The inhibition of Fz receptors by analogues of either Wnt3a/Wnt5A or sfrp2 leads to a reduction in the extent of the MI and development of subsequent heart insufficiency (Ref. 98).

Furthermore, Wnt signalling is crucial for the loosening of AJs in an endothelial monolayer, facilitating the infiltration of leukocytes into the vessel wall. AJs are formed by interactions of transmembrane proteins, namely, VE-cad, of neighbouring cells. The cytosolic domains of these proteins are anchored to the actin cytoskeleton by a number of proteins, the most important of which are p120, plakoglobin, α -catenin and β -cat (Ref. 99). Tyrosine phosphorylation within the cytoplasmic domain of VE-cadherin causes dissociation of catenins and destabilisation of AJs with a subsequent rise in endothelial permeability (Ref. 100). A side effect of such dissociation is a higher accumulation of β -cat in the cytosol available for translocation to the nucleus and activation of target genes.

Haidari *et al.* demonstrated that exposure of ECs to hyperglycaemic conditions results in tyrosine phosphorylation of VE-cadherin, dissociation of β -cat from AJs, and enhanced transendothelial migration (Ref. 67). Notably, this process was mediated by PKC- β , a downstream effector in the β -cateninindependent Wnt signalling pathway. Moreover, a high concentration of glucose led to PKC-mediated GSK3 β phosphorylation with a subsequent rise in the cytosolic pool of β -cat and activation of the β -cat/TCF/Lef-1 responsive promoter. Hyperglycaemia significantly increased the expression of β -catenin target genes such as cycD1 (cyclin D1) and u-PA (urokinase), indicating that exposure to high glucose levels activates the β -catenin-dependent Wnt pathway in ECs. These findings may provide an important link between diabetes and accelerated atherogenesis (Ref. 76).

Another study reported that VEGF stimulates uPA expression by inducing EC hyperpermeability through β -cat-dependent urokinase-type plasminogen activator receptor (uPA/uPAR) activation (Refs 101, 102). These results suggest that the crosstalk of the TGF- β and Wnt pathways is important in endothelial dysfunction and that the u-PA/uPAR and Wnt pathways might provide a molecular bridge linking the inflammation and progression of atherosclerosis in patients with diabetes. Additionally, hyperglycaemia promotes a pro-coagulant phenotype in ECs by upregulating the expression of plasminogen activator inhibitor (PAI-1) and P selectin (P-Sel) as well as inducing the secretion of fibrinogen (Ref. 103).

TUG1 is a long non-coding RNA (lncRNA) that is highly expressed in ECs (Ref. 104). TUG1 overexpression was detected in CAD tissues compared with normal arterial tissues. Upregulation of TUG1 was observed in high-dose glucose-induced HUVECs. This overexpression of TUG1 stimulated a number of genes in HUVECs: cell cycle-related, proliferation-related and Wnt pathway-related (e.g., β -catenin, c-myc). The Wnt pathway might be associated with TUG1-promoted migration and proliferation of ECs. This could be a possible mechanism of TUG1 involvement in the regulation of atherosclerosis in DM (Ref. 68).

DKK-1, an inhibitor of β -catenin-dependent Wnt signalling, has been observed at higher concentrations in clinical and experimental atherosclerosis compared with normal tissue. The main sources of DKK-1 are ECs and platelets (Ref. 105). The plasma concentration of DKK-1 was higher in patients with T2DM compared with healthy controls, which was associated with platelet activation markers and increased levels of endothelial dysfunction. Additionally, improved glycaemic control downregulates DKK-1 expression in T2DM (Ref. 106).

Wnt in diabetic retinopathy (DR)

The key factors underlying the development of DR are inflammation (mediated by VEGF, ICAM and TNF α), microvascular damage and oxidative stress, which lead to a disruption of the bloodretina barrier and retinal ischaemia. Subsequent retinal vascular leakage and neovascularisation result in loss of sight (Refs 107, 108). Since VEGF, TNF α and ICAM are target genes of the Wnt signalling pathway, this pathway has become an important field of research in the pathophysiology of DR (Ref. 109). The β -catenin-dependent Wnt pathway has been shown to be upregulated in both human and rodent models of DR. Using a murine model, Zhou *et al.* (Ref. 110) showed that under ischaemic conditions, as well as in induced diabetes, β -catenin knockout in Muller retinal cells reduces neovascularisation, vascular leakage and inflammation in comparison with those in wild-type mice. At the molecular level, the lack of β -cat reduced the production of VEGF and TNF α . These results further support the detrimental effect of the Wnt pathway in the pathogenesis of DR. Additionally, pericyte loss, which is an important hallmark of DR, was significantly lower in β -cat knockout mice than in wild-type mice exposed to the same hyperglycaemic conditions (Ref. 87). Furthermore, Lee et al. (Ref. 68) demonstrated that application of an anti-LRP6 monoclonal antibody on retinal ECs led to reduced β -cat accumulation, lower expression of VEGF5 and TNF α induced by hyperglycaemia in vitro. Furthermore, an intravitreal administration of the same antibody decreased vascular leakage and neovascularisation in a rat DR model (Ref. 68). Similarly, DKK1, a natural inhibitor of the Wnt signalling pathway, has been shown to alleviate retinal inflammation and vascular leakage in DR (Refs 69, 88). Another negative regulator of Wnt/ β -cat signalling, Adenomatosis Polyposis Coli Downregulated 1 Protein (Apcdd1), is expressed in retinal ECs during angiogenesis and barrier formation. Apcdd1-deficient mice exhibit a transient increase in vessel density. Moreover, mice that overexpress Apcdd1 in retina ECs have reduced vessel density but increased barrier permeability (Ref. 111).

Another study focusing on fibrosis in DR has shown that introducing SERPINA3 K, a serine proteinase inhibitor, reduces fibrosis in a rat DR model by antagonizing connective tissue growth factor (CTGF), a potent fibrogenic factor, and minimizing the production of ECM proteins. Thus, the effect of SERPINA3 K is likely to be achieved *via* the inhibition of the Wnt signalling pathway (Ref. 112).

Adherens junctions and the β -catenin-dependent Wnt pathway

In ECs, β -catenin performs several functions owing to its ability to bind different molecular partners (Fig. 2). One of them is the formation and stabilisation of tight AJs in association with the extracellular membrane protein E-cadherin (E-cad). The central part of the β -cat amino acid chain contains binding sites for E-cad (a crucial protein for cell-cell interactions), LEF-1 (a transcription factor) and APC (a part of the β -catenin degrading complex). As all these binding sites overlap, β -cat can interact only with one of these partners at a given time (Refs 113, 114). Thus, only upon dissociation from E-cad does β -cat become available for the other two proteins and can function as the Wnt pathway component. This phenomenon has already been demonstrated in Xenopus (Ref. 115) and Drosophila (Ref. 116). Moreover, β -cat released from AJs upon their dissociation forms a pool from which it is transported to the nucleus upon activation of the Wnt pathway (Ref. 117). Loss of E-cad expression has been reported to result in β -cat accumulation in the cytoplasm, with its subsequent nuclear translocation and interaction with LEF (Ref. 118). These observations confirm the involvement of the β -catenin-dependent Wnt pathway in the disintegration of the EC monolayer, leading to the loss of one of its basic roles in maintaining vessel homeostasis.

Research in progress and outstanding research questions

Chronic hyperglycaemia extensively influences the vasculature, leading to multiple organ complications in patients suffering from DM. As DM is a growing problem worldwide, there is and will be more demand for new therapies to treat these complications. Previous studies indicate the possible involvement of Wnt signalling in the development of hyperglycaemia-related complications such as atherosclerosis, DR and certain types of cancers. Given the importance of hyperglycaemia in disrupting physiological cellular processes and leading to multiple complications, it is crucial to remember that normalizing blood glucose concentration remains the basic course of therapy in diabetes-related health issues. One of the most commonly prescribed antihyperglycaemia drugs, metformin, has been shown to reduce the risk and slow the progression of several types of cancer. Many mechanisms have been proposed as an explanation of this phenomenon, the most simple of which may be reducing hyperglycaemia and its cellular consequences (Ref. 119). However, there are elements of the pathogenesis of DM complications that are irreversible, such as metabolic memory, which requires a more complex approach and further investigation into potential therapies.

The current pharmacotherapy for slowing the process of vascular complications in patients with diabetes is rather narrow and is mostly focused on lowering LDL cholesterol levels. Most likely, the most widely used pharmacological agents are statins, which in addition to lowering LDL, have been shown to stabilise the atherosclerotic plaque, preventing thrombus formation. These, however, are not always effective and have a number of well-characterised adverse effects, such as liver dysfunction or rhabdomyolysis, which may lead to acute kidney injury. Similar reservations apply to fibrates, which also reduce lipid levels and predispose them to nearly the same adverse effects. An emerging new player in the field is the anti-PCSK-9 antibody, evolocumab, which targets and inactivates LDL cholesterol receptors. In light of the presented data, the effect of evolocumab on Wnt signalling in diabetic vascular complications appears to be a very interesting alternative to the current use of statins.

One of the most important vascular complications in diabetes is fibrosis, which is directly related to EndoMT. ECs are progenitors of cardiac pericytes and vascular smooth muscle cells, both of which are involved in tissue regeneration and vascular remodelling (Refs 40–42). Blocking Wnt signalling with a homologous peptide fragment of wnt3a/wnt5a has been reported to reduce infarct expansion and prevent heart failure after MI (Ref. 98). Additionally, the occurrence of infarction-related fibrosis can be reduced by a small molecule allosteric activator of aldehyde dehydrogenase-2 (ALDH2), which downregulates β -cat, phosphorylated GSK-3 β and Wnt-1 (Ref. 120). Based on the aforementioned studies, inhibition of Wnt signalling can be considered to reduce vascular complications, both in patients with diabetes and non-diabetic individuals.

Untreated ocular complications of chronic hyperglycaemia lead to proliferative DR, diabetic macular oedema and eventually loss of sight. Several sight-saving therapies, such as laser photocoagulation or vitrectomy, are available; however, none of them is fully satisfactory, and there is a need for the development of new therapies in this area (Ref. 4). Among the new therapeutic candidates are anti-VEGF agents (Refs 82, 83). Although some promising results have been presented for these candidates, this therapy is not effective for all patients. As described above, the β -catenin-dependent Wnt pathway appears to be involved in many pathological processes, such as neoangiogenesis or inflammation, resulting in VEGF, $TNF\alpha$ and ICAM expression and eventually leading to DR. As such, this signalling system could be a potential target for alleviating inflammation and neoangiogenesis. Thus far, the anti-LRP6 monoclonal antibody is the only potential the rapeutic approach targeting the β -cat-dependent system proposed to treat DR (Ref. 68). Another potential therapeutic target could be Apcdd1 or ALDH2, overexpression or over-activity of which downregulates Wnt-dependent genes (Refs 111, 120).

Finally, disrupting the Wnt pathway could be considered a potential approach to limit the development of certain types of

cancers that are more frequent in patients with chronic hyperglycaemia.

OMP-18R5, a monoclonal antibody interacting with five Fzd receptors, inhibited different human tumour growth in xenografts, diminished tumour-initiating cell frequency and exhibited synergistic activity with standard chemotherapeutic agents (Ref. 121). NSC654259, another small molecule that targets the cysteine-rich domain of Frizzled, has shown attenuating properties in multiple types of tumour cell lines (Ref. 122).

An eye-specific miRNA-184 expressed in the lens, cornea and retina (Ref. 123) was significantly downregulated in a murine model of oxygen-induced retinopathy (Ref. 124). Bioinformatics analysis showed that it targets several components of Wnt signalling such as Wnt ligands (Wnt-9, Wnt-16) and Wnt receptors (Frizzled-3, Frizzled-7) (Refs 109, 125), showing therapeutic potential in proliferative retinopathy.

Increased levels of LRP6 have been observed in the retina of DR animal models (Ref. 109). Mab2F1, a monoclonal antibody against the E1E2 domain of LRP6, inhibited Wnt signalling upregulation caused by high glucose levels and suppressed the expression of inflammatory factors (VEGF, ICAM-1 and TNF-a) (Ref. 68).

In summary, there is growing evidence that abnormalities at different stages of Wnt signalling result in inflammation, neoangiogenesis and abnormal gene expression. All these phenomena underlie a series of complications in diabetes. Regulating these processes, for example, by inhibiting the Fz or LRP receptors or downregulating the Wnt β -catenin-dependent pathway, could lead to the development of a multipotent therapy, improving the condition of patients suffering from diabetic complications.

Summary

Many diseases that are associated with EC dysfunction are related to an impaired Wnt pathway. Accumulating evidence confirms the idea that in hyperglycaemic conditions, the endothelium loses its protective abilities, becoming activated (inflamed) and undergoing EndoMT, enhancing the expression of Wnt-dependent genes involved in cell proliferation and permeability. In these processes, β -cat signalling appears to be the most important, showing the need for further investigation to establish whether Wnt pathway inhibitors or antagonists may have clinical relevance in the treatment of vascular diabetic complications such as accelerated atherosclerosis and its consequences, DR or DN.

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Highlights

- Diabetes favours oncogenesis and accelerates cardiovascular complications.
- Hyperglycaemia affects endothelial functions and induces inflammatory processes leading to atherosclerosis and impaired angiogenesis.
- Wnt signalling is activated during hyperglycaemia and causes endothelial dysfunction and EndoMT.
- Targeting Wnt pathways improves hyperglycaemia-related endothelial complications.

References

- 1. vanHinsbergh VWM (2012) Endothelium role in regulation of coagulation and inflammation. *Seminars in Immunopathology* **34**, 93–106.
- Palmer RM et al. (1987) Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. Nature 327, 524–526.

- 3. Montoro-García S *et al.* (2014) Potential value of targeting vonWillebrand factor in atherosclerotic cardiovascular disease. *Expert Opinion on Therapeutic Targets* 18, 43–53.
- 4. Stegenga ME et al. (2006) Hyperglycemia stimulates coagulation, whereas hyperinsulinemia impairs fibrinolysis in healthy humans. *Diabetes* 55, 1807–1812.
- Dogné S et al. (2018) Endothelial glycocalyx as a shield against diabetic vascular complications: involvement of hyaluronan and hyaluronidases. Arteriosclerosis, Thrombosis, and Vascular Biology 38, 1427– 1439.
- Popov D (2010) Endothelial cell dysfunction in hyperglycemia: phenotypic change, intracellular signaling modification, ultrastructural alteration, and potential clinical outcomes. *International Journal of Diabetes and Clinical Research* 2, 189–195.
- Laughlin MH et al. (2008) Importance of hemodynamic forces as signals for exercise-induced changes in endothelial cell phenotype. *Journal of Applied Physiology* 104, 588–600.
- 8. Cade W (2008) Diabetes-related microvascular and macrovascular diseases in the physical therapy setting. *Physical Therapy* 88, 1322–1335.
- 9. Funamoto K et al. (2017) Endothelial monolayer permeability under controlled oxygen tension. *Integrative Biology* 9, 529–538.
- Hirose A et al. (2010) Advanced glycation end products increase endothelial permeability through the RAGE/Rho signaling pathway. FEBS Letters 584, 61–66.
- Rho SS et al. (2017) Dynamic regulation of vascular permeability by vascular endothelial cadherin-mediated endothelial cell-cell junction. *Journal of Nippon Medical School* 84, 148–159.
- Fraisl P et al. (2009) Regulation of angiogenesis by oxygen and metabolism. Developmental Cell 16, 167–179.
- Mundi S et al. (2018) Endothelial permeability, LDL, deposition, and cardiovascular risk factors-a review. Cardiovascular Research 114, 34–52.
- 14. Avogadro A et al. (2011) Endothelial dysfunction in diabetes: the role of reparatory mechanisms. *Diabetes Care* 34, 285–290.
- Brunssen C et al. (2010) COUP-TFIIis regulated by high glucose in endothelial cells. Hormone and Metabolic Research 42, 189–195.
- Williams SB et al. (1998) Acute hyperglycemia attenuates endotheliumdependent vasodilation in humans in vivo. Circulation 97, 1695–1701.
- David L et al. (2008) Bone morphogenetic protein-9 is a vascular quiescence factor. Circulation Research 102, 914–922.
- Tanaka A et al. (2010) Inhibition of endothelial cell activation by bHLH protein E2-2 and its impairment of angiogenesis. Blood 115, 4138–4147.
- Singh H et al. (2010) High glucose and elevated fatty acids suppress signaling by the endothelium protective ligand angiopoietin-1. *Microvascular Research* 79, 121–127.
- Fukuhara S et al. (2010) Angiopoietin-1/Tie2 receptor signaling in vascular quiescence and angiogenesis. Histology & Histopathology 25, 387–396.
- Jiang A et al. (2009) Loss of VLDL receptor activates retinal vascular endothelial cells and promotes angiogenesis. *Investigative Ophthalmology & Visual Science* 50, 844–850.
- 22. Xu L et al. (2009) Promoter cloning and characterization of the antivascular proliferation gene R-ras. Role of ETS- and SP-binding motifs. *Journal of Biological Chemistry* 284, 2706–2718.
- 23. Rajapakse AG et al. (2009) The hexosamine biosynthesis inhibitor azaserine prevents endothelial inflammation and dysfunction under hyperglycemic condition through antioxidant effects. The American Journal of Physiology-Heart and Circulatory Physiology 296, 815–822.
- Liu Y et al. (2009) Effect of diabetes duration and high glucose on retinal interleukin-1b expression. *Investigative Ophthalmology & Visual Science* 50, 4984. E-Abstract.
- 25. Bhatwadekar AD *et al.* (2009) Retinal endothelial cell apoptosis stimulates recruitment of endothelial progenitor cells. *Investigative Ophthalmology & Visual Science* **50**, 4967–4973.
- Shakhov AS et al. (2014) Reorganization of endothelial cells cytoskeleton during formation of functional monolayer in vitro. Cell and Tissue Biology 8, 138–151.
- Targosz-Korecka M et al. (2013) Stiffness memory of EA.hy926 endothelial cells in response to chronic hyperglycemia. Cardiovascular Diabetology 12, 96.
- Sharma A et al. (2017) The nuclear factor (erythroid-derived 2)-like 2 (Nrf2) activator dh404 protects against diabetes-induced endothelial dysfunction. *Cardiovascular Diabetology* 16, 33.

- Bucciarelli LG et al. (2009) Inflammatory stress in primary venous and aortic endothelial cells of type 1 diabetic mice. Diabetes & Vascular Disease Research 6, 249–261.
- Popov D et al. (2006) Cellular mechanisms and signalling pathways activated by high glucose and AGE-albumin in the aortic endothelium. Archives of Physiology and Biochemistry 112, 265–273.
- Trudeau K et al. (2009) High glucose alters mitochondrial morphology and membrane potential heterogeneity in retinal endothelial cells. Investigative Ophthalmology & Visual Science 50, 33. E-Abstract.
- 32. Dokken BB (2008) The pathophysiology of cardiovascular disease and diabetes: beyond blood pressure and lipids. *Diabetes Spectrum* 21, 160–165.
- 33. Klaassen I et al. (2013) Molecular basis of the inner blood-retinal barrier and its breakdown in diabetic macular edema and other pathological conditions. *Progress in Retinal and Eye Research* 34, 19–48.
- 34. Naka K et al. (2012) Determinants of vascular function in patients with type 2 diabetes. Cardiovascular Diabetology 11, 127.
- Reddy MA et al. (2015) Epigenetic mechanisms in diabetic complications and metabolic memory. Diabetologia 58, 443–455.
- Sánchez-Duffhues G et al. (2018) Endothelial-to-mesenchymal transition in cardiovascular diseases: developmental signaling pathways gone awry. Developmental Dynamics 247, 492–508.
- Van Meeteren LA et al. (2012) Regulation of endothelial cell plasticity by TGF-β. Cell and Tissue Research 347, 177–186.
- Chen Q et al. (2016) Endothelial cells are progenitors of cardiac pericytes and vascular smooth muscle cells. *Nature Communications* 7, 12422.
- Li J et al. (2009) Endothelial-myofibroblast transition contributes to the early development of diabetic renal interstitial fibrosis in streptozotocininduced diabetic mice. *The American Journal of Pathology* 175, 1380– 1388.
- Widyantoro B et al. (2010) Endothelial cell-derived endothelin-1 promotes cardiac fibrosis in diabetic hearts through stimulation of endothelial-to-mesenchymal transition. *Circulation* 175, 1380–1388.
- Aronson D et al. (2002) How hyperglycemia promotes atherosclerosis: molecular mechanisms. Cardiovascular Diabetology 1, 1–10.
- 42. Wu X et al. (2017) Exosomes from high glucose-treated glomerular endothelial cells trigger the epithelial-mesenchymal transition and dys-function of podocytes. *Scientific Reports* 7, 9371.
- Korinek V et al. (1998) Two members of the Tcf family implicated in Wnt/beta-catenin signaling during embryogenesis in the mouse. Molecular and Cellular Biology 18, 1248–1256.
- 44. Lowry WE et al. (2005) Defining the impact of beta-catenin/Tcf transactivation on epithelial stem cells. Genes & Development 19, 1596–1611.
- 45. Fahiminiya S et al. (2013) Mutations in WNT1 are a cause of osteogenesis imperfecta. *Journal of Medical Genetics* 50, 345–348.
- Chen AE et al. (2005) Protein kinase A signalling via CREB controls myogenesis induced by Wnt proteins. *Nature* 433, 317–322.
- 47. Kanazawa A *et al.* (2004) Association of the gene encoding wingless-type mammary tumor virus integration-site family member 5B (WNT5B) with type 2 diabetes. *American Journal of Human Genetics* **75**, 832–843.
- Christodoulides C et al. (2006) WNT10B mutations in human obesity. Diabetologia 49, 678–684.
- 49. Habas R et al. (2005) Dishevelled and Wnt signaling: is the nucleus the final frontier? The Journal of Biology 4, 2.
- 50. **Dessimoz J** *et al.* (2005) Pancreas-specific deletion of β -catenin reveals Wnt-dependent and Wnt-independent functions during development. *Current Biology* **15**, 1677–1683.
- Schinner S et al. (2008) Regulation of insulin secretion, glucokinase gene transcription and beta cell proliferation by adipocyte-derived Wnt signalling molecules. *Diabetologia* 51, 147–154.
- 52. **Rulifson IC** *et al.* (2007) Wnt signaling regulates pancreatic β cell proliferation. *Proceedings of the National Academy of Sciences of the USA* **104**, 6247–6252.
- Liu Z et al. (2009) Stromal cell-derived factor-1 promotes survival of pancreatic beta cells by the stabilization of beta-catenin and activation of transcription factor 7-like 2 (TCF7L2). Diabetologia 52, 1589–1598.
- 54. **Fujino T** *et al.* (2003) Low-density lipoprotein receptor-related protein 5 (LRP5) is essential for normal cholesterol metabolism and glucose-induced insulin secretion. *Proceedings of the National Academy of Sciences of the USA* **100**, 229–234.

- Liu Z et al. (2008) Glucagon-like peptide-1 activation of TCF7L2-dependent Wnt signaling enhances pancreatic b cell proliferation. *Journal of Biological Chemistry* 283, 8723–8735.
- Cheng CW et al. (2003) Wnt-1 signaling inhibits human umbilical vein endothelial cell proliferation and alters cell morphology. *Experimental Cell Research* 291, 415–425.
- 57. Choi HJ et al. (2012) The Wnt pathway and the roles for its antagonists, Dkks, in angiogenesis. *Life* 64, 724–731.
- Mao C et al. (2000) Differential expression of rat frizzled-related frzb-1 and frizzled receptor fz1 and fz2 genes in the rat aorta after balloon injury. Arteriosclerosis, Thrombosis, and Vascular Biology 20, 43–51.
- 59. Goodwin AM et al. (2006) Cultured endothelial cells display endogenous activation of the canonical Wnt signaling pathway and express multiple ligands, receptors, and secreted modulators of Wnt signaling. *Developmental Dynamics* 235, 3110–3120.
- Anagnostou SH *et al.* (2008) Glucose induces an autocrine activation of the Wnt/β-catenin pathway in macrophage cell lines. *Biochemical Journal* 416, 211–218.
- Komiya Y et al. (2008) Wnt signal transduction pathways. Organogenesis 4, 68–75.
- 62. Minde DP et al. (2011) Messing up disorder: how do missense mutations in the tumor suppressor protein APC lead to cancer? *Molecular Cancer* 10, 101.
- Chandramouli A et al. (2013) Gli activity is critical at multiple stages of embryonic mammary and nipple development. PLoS ONE 8, e79845.
- 64. Jho E *et al.* (2002) Wnt/beta-catenin/Tcf signaling induces the transcription of Axin2, a negative regulator of the signaling pathway. *Molecular and Cellular Biology* **22**, 1172–1183.
- 65. Christodoulides C et al. (2006) The Wnt antagonist Dickkopf-1 and its receptors are coordinately regulated during early human adipogenesis. *Journal of Cell Science* 119, 2613–2620.
- 66. Cawthorn WP et al. (2007) Tumour necrosis factor-alpha inhibits adipogenesis via a beta-catenin/TCF4(TCF7L2)-dependent pathway. Cell Death & Differentiation 14, 1361–1373.
- Haidari M et al. (2014) Disruption of endothelial adherens junctions by high glucose is mediated by protein kinase C-β-dependent vascular endothelial cadherin tyrosine phosphorylation. *Cardiovascular Diabetology* 13, 105.
- Lee K et al. (2012) Therapeutic potential of a monoclonal antibody blocking the Wnt pathway in diabetic retinopathy. *Diabetes* 61, 2948– 2957.
- Holnthoner W et al. (2002) Fibroblast growth factor-2 induces Lef/ Tcf-dependent transcription in human endothelial cells. Journal of Biological Chemistry 277, 45847–45853.
- Gelfand BD et al. (2011) Hemodynamic activation of beta-catenin and T-cell-specific transcription factor signaling in vascular endothelium regulates fibronectin expression. Arteriosclerosis, Thrombosis, and Vascular Biology 31, 1625–1633.
- Hou N et al. (2016) Transcription factor 7-like 2 mediates canonical Wnt/ β-Catenin signaling and c-myc upregulation in heart failure. *Circulation: Heart Failure* 9, 1–18. doi: 10.1161/CIRCHEARTFAILURE.116.003010 e003010.
- Cesare AJ et al. (2010) Alternative lengthening of telomeres. Models, mechanisms, and implications. *Nature Reviews Genetics* 11, 319–330.
- Bilsland AE et al. (2006) Transcriptional repression of telomerase RNA gene expression by c-Jun-NH2-kinase and Sp1/Sp3. Cancer Research 66, 1363–1370.
- 74. **Deng WG** *et al.* (2007) Tumor-specific activation of human telomerase reverses transcriptase promoter activity by activating enhancer-binding protein- 2β in human lung cancer cells. *Journal of Biological Chemistry* **282**, 26460–26470.
- 75. Zhang Y et al. (2012) Human Telomerase Reverse Transcriptase (hTERT) is a novel target of the Wnt/β-catenin pathway in human cancer. Journal of Biological Chemistry 289, 32494–32511.
- Park JI et al. (2009) Telomerase modulates Wnt signalling by association with target gene chromatin. *Nature* 460, 66–72.
- Khandekar MJ et al. (2011) Molecular mechanisms of cancer development in obesity. Nature Reviews Cancer 11, 886–895.
- Chocarro-Calvo A et al. (2013) Glucose-induced β-catenin acetylation enhances Wnt signaling in cancer. Molecular Cell 49, 474–486.
- 79. Chouhan S et al. (2016) Glucose induced activation of canonical Wnt signaling pathway in hepatocellular carcinoma is regulated by DKK4. *Scientific Reports* 6, 27558.

- Coughlin SS *et al.* (2004) Diabetes mellitus as a predictor of cancer mortality in a large cohort of US adults. *American Journal of Epidemiology* 159, 1160–1167.
- Jee SH et al. (2005) Fasting serum glucose level and cancer risk in Korean men and women. Journal of the American Medical Association 293, 194–202.
- Baron JA et al. (2001) Metabolic disorders and breast cancer risk (United States). Cancer Causes and Control 12, 875–880.
- Lipscombe LL et al. (2006) Diabetes mellitus and breast cancer: a retrospective population-based cohort study. Breast Cancer Research and Treatment 98, 349–356.
- Wideroff L et al. (1997) Cancer incidence in a population-based cohort of patients hospitalized with diabetes mellitus in Denmark. *Journal of the National Cancer Institute* 89, 1360–1365.
- Friberg E et al. (2007) Diabetes mellitus and risk of endometrial cancer: a meta-analysis. Diabetologia 50, 1365–1374.
- Anderson KE et al. (2001) Diabetes and endometrial cancer in the Iowa Women's Health Study. *Cancer Epidemiology Biomarkers and Prevention* 10, 611–616.
- Vigneri P et al. (2009) Diabetes and cancer. Endocrine-Related Cancer 16, 1103–1123.
- Yun J et al. (2009) Glucose deprivation contributes to the development of KRAS pathway mutations in tumor cells. Science 325, 1555–1559.
- Tseng CH et al. (2009) Age-related risk of mortality from bladder cancer in diabetic patients: a 12-year follow-up of a national cohort in Taiwan. *Annals of Medicine* 41, 371–379.
- Lewis JD et al. (2011) Risk of bladder cancer among diabetic patients treated with pioglitazone: interim report of a longitudinal cohort study. *Diabetes Care* 34, 916–922.
- Masur K et al. (2011) Diabetogenic glucose and insulin concentrations modulate transcriptom and protein levels involved in tumour cell migration, adhesion and proliferation. British Journal of Cancer 104, 345–352.
- Harding JL et al. (2015) Cancer risk among people with type 1 and type 2 diabetes: disentangling true associations, detection bias, and reverse causation. *Diabetes Care* 38, 264–270.
- Yan HY et al. (2018) TUG1 promotes diabetic atherosclerosis by regulating proliferation of endothelial cells via Wnt pathway. The European Review for Medical and Pharmacological 22, 6922–6929.
- Kim J et al. (2010) Wnt5a induces endothelial inflammation via betacatenin-independent signaling. Journal of Immunology 185, 1274–1282.
- Christman MA et al. (2008) Wnt5a is expressed in murine and human atherosclerotic lesions. American Journal of Physiology. Heart and Circulatory Physiology 294, 2864–2870.
- Bhatt PM et al. (2012) Increased Wnt5a mRNA expression in advanced atherosclerotic lesions, and oxidized LDL treated human monocytederived macrophages. The Open Circulation & Vascular Journal 5, 1–7.
- Malgor R et al. (2014) Wnt5a, TLR2 and TLR4 are elevated in advanced human atherosclerotic lesions. *Inflammation Research* 63, 277–285.
- Laeremans H et al. (2011) Blocking of frizzled signaling with a homologous peptide fragment of wnt3a/wnt5a reduces infarct expansion and prevents the development of heart failure after myocardial infarction. *Circulation* 124, 1626–1635.
- Dejana E et al. (2008) The role of adherens junctions and VE-cadherin in the control of vascular permeability. *Journal of Cell Science* 121, 2115– 2122.
- Schulte D et al. (2011) Stabilizing the VE-cadherin-catenin complex blocks leukocyte extravasation and vascular permeability. EMBO Journal 30, 4157–4170.
- Behzadian MA et al. (2003) VEGF-induced paracellular permeability in cultured endothelial cells involves urokinase and its receptor. FASEB Journal 17, 753–754.
- Esser S et al. (1998) Vascular endothelial growth factor induces VE-cadherin tyrosine phosphorylation in endothelial cells. *Journal of Cell Science* 111, 1853–1865.
- 103. Goldberg RB (2009) Cytokine and cytokine-like inflammation markers, endothelial dysfunction, and imbalanced coagulation in development of diabetes and its complications. *The Journal of Clinical Endocrinology* and Metabolism 94, 3171–3182.

- 104. Chen C et al. (2016) Tanshinol suppresses endothelial cells apoptosis in mice with atherosclerosis via lncRNA TUG1up-regulating the expression of miR-26a. American Journal of Translational Research 8, 2981–2991.
- Ueland T et al. (2009) Dickkopf-1 enhances inflammatory interaction between platelets and endothelial cells and shows increased expression in atherosclerosis. Arteriosclerosis Thrombosis and Vascular Biology 29, 1228–1234.
- 106. Lattanzio S et al. (2014) Circulating dickkopf-1 in diabetes mellitus: association with platelet activation and effects of improved metabolic control and low-dose aspirin. Journal of the American Heart Association 3, 1–10. doi: 10.1161/JAHA.114.001000.
- 107. Klein BEK et al. (2009) The relation of markers of inflammation and endothelial dysfunction to the prevalence and progression of diabetic retinopathy: Wisconsin epidemiologic study of diabetic retinopathy. *Archives of Ophthalmology* 127, 1175–1182.
- van Hecke MV et al. (2005) Inflammation and endothelial dysfunction are associated with retinopathy: the Hoorn Study. *Diabetologia* 48, 1300–1306.
- 109. Chen Y et al. (2009) Activation of the Wnt pathway plays a pathogenic role in diabetic retinopathy in humans and animal models. *The American Journal of Pathology* 175, 2676–2685.
- Zhou KK et al. (2014) Interruption of Wnt signaling in Müller cells ameliorates ischemia-induced retinal neovascularization. PLoS ONE 9, e108454.
- Mazzoni J et al. (2017) The Wnt inhibitor Apcdd1 coordinates vascular remodeling and barrier maturation of retinal blood vessels. *Neuron* 96, 1055–1069.
- 112. Zhang B et al. (2010) Inhibition of connective tissue growth factor overexpression in diabetic retinopathy by SERPINA3 K via blocking the WNT/beta-catenin pathway. *Diabetes* 59, 1809–1816.
- Hülsken J et al. (1994) E-cadherin and APC compete for the interaction with beta-catenin and the cytoskeleton. *Journal of Cell Biology* 127, 2061– 2069.
- Orsulic S et al. (1999) E-cadherin binding prevents beta-catenin nuclear localization and beta-catenin/LEF-1-mediated transactivation. *Journal of Cell Science* 112, 1237–1245.
- 115. McMahon AP et al. (1989) Ectopic expression of the proto-oncogene int-1 in Xenopus embryos leads to duplication of the embryonic axis. *Cell* 58, 1075–1084.
- Cox RT et al. (1996) Armadillo is required for adherens junction assembly, cell polarity, and morphogenesis during Drosophila embryogenesis. Journal of Cell Biology 134, 133–148.
- 117. Kam Y et al. (2009) Cadherin-bound beta-catenin feeds into the Wnt pathway upon adherens junctions dissociation: evidence for an intersection between beta-catenin pools. PLoS ONE 4, e4580.
- 118. Eger A et al. (2000) Epithelial mesenchymal transition by c-Fos estrogen receptor activation involves nuclear translocation of beta-catenin and upregulation of beta-catenin/lymphoid enhancer binding factor-1 transcriptional activity. Journal of Cell Biology 148, 173–188.
- Aljada A et al. (2012) Metformin and neoplasia: implications and indications. Pharmacology & Therapeutics 133, 108–115.
- 120. Zhao X et al. (2015) Aldehyde dehydrogenase-2 protects against myocardial infarction-related cardiac fibrosis through modulation of the Wnt/β-catenin signaling pathway. Therapeutics and Clinical Risk Management 11, 1371–1381.
- 121. Gurney A et al. (2012) Wnt pathway inhibition via the targeting of frizzled receptors results in decreased growth and tumorigenicity of human tumors. PNAS 109, 11717–11722.
- 122. Lee HJ et al. (2015) Structure-based discovery of novel small molecule Wnt signaling inhibitors by targeting the cysteine-rich domain of frizzled. Journal of Biological Chemistry 290, 30596–30606.
- Ryan DG et al. (2006) MicroRNAs of the mammalian eye display distinct and overlapping tissue specificity. *Molecular Vision* 12, 1175–1184.
- 124. Takahashi Y et al. (2015) MicroRNA-184 modulates canonical Wnt signaling through the regulation of frizzled-7 expression in the retina with ischemia-induced neovascularization. FEBS Letters 589, 1143–1149.
- 125. Shen J et al. (2008) MicroRNAs regulate ocular neovascularization. Molecular Therapy 16, 1208–1216.